

ExomeDepth

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Contents

1	What ExomeDepth does and tips for QC	1
1.1	What ExomeDepth does and does not do	1
1.2	Useful quality checks	1
2	Create count data from BAM files	1
2.1	Count for the autosomes	1
2.2	Counts for chromosome X	2
3	Load an example dataset	3
4	Build the most appropriate reference set	3
5	CNV calling	4
6	Better annotation of CNV calls	4
7	A visual example	6
8	How to loop over the multiple samples	7
9	Technical information about R session	8

1 What ExomeDepth does and tips for QC

1.1 What ExomeDepth does and does not do

ExomeDepth uses read depth data to call CNVs from exome sequencing experiments. A key idea is that the test exome should be compared to a matched aggregate reference set. This aggregate reference set should combine exomes from the same batch and it should also be optimized for each exome. It will certainly differ from one exome to the next.

Importantly, ExomeDepth assumes that the CNV of interest is absent from the aggregate reference set. Hence related individuals should be excluded from the aggregate reference. It also means that ExomeDepth can miss common CNVs, if the call is also present in the aggregate reference. ExomeDepth is really suited to detect rare CNV calls (typically for rare Mendelian disorder analysis).

The ideas used in this package are of course not specific to exome sequencing and could be applied to other targeted sequencing datasets, as long as they contain a sufficiently large number of exons to estimate the parameters (at least 20 genes, say, but probably more would be useful). Also note that PCR based enrichment studies are often not well suited for this type of read depth analysis. The reason is that as the number of cycles is often set to a high number in order to equalize the representation of each amplicon, which can discard the CNV information.

1.2 Useful quality checks

Just to give some general expectations I usually obtain 150-280 CNV calls per exome sample (two third of them deletions). Any number clearly outside of this range is suspicious and suggests that either the model was inappropriate or that something went wrong while running the code. Less important and less precise, I also expect the

aggregate reference to contain 5-10 exome samples. While there is no set rule for this number, and the code may work very well with fewer exomes in the aggregate reference set, numbers outside of this range suggest potential technical artifacts.

2 Create count data from BAM files

2.1 Count for the autosomes

Firstly, to facilitate the generation of read count data, exon positions for the hg19 build of the human genome are available within `ExomeDepth`. This `exons.hg19` data frame can be directly passed as an argument of `getBAMCounts` (see below).

```
> library(ExomeDepth)
> data(exons.hg19)
> print(head(exons.hg19))
```

	chromosome	start	end	name
1	1	12011	12058	DDX11L10-201_1
2	1	12180	12228	DDX11L10-201_2
3	1	12614	12698	DDX11L10-201_3
4	1	12976	13053	DDX11L10-201_4
5	1	13222	13375	DDX11L10-201_5
6	1	13454	13671	DDX11L10-201_6

To generate read count data, the function `getBamCounts` in `ExomeDepth` is set up to parse the BAM files. It generates an array of read count, stored in a `GenomicRanges` object. It is a wrapper around the function `countBamInGRanges.exomeDepth` which is derived from an equivalent function in the `exomeCopy` package. You can refer to the help page of `getBAMCounts` to obtain the full list of options. An example line of code (not evaluated here) would look like this:

```
> data(exons.hg19)
> my.counts <- getBamCounts(bed.frame = exons.hg19,
+                           bam.files = my.bam,
+                           referenceFasta = fasta)
```

`my.bam` is a set character vector of indexed BAM files. `fasta` is the reference genome in fasta format (only useful if one wants to obtain the GC content). `exons.hg19` are the positions and names of the exons on the hg19 reference genome (as shown above).

`getBAMCounts` creates an object of the `GRanges` class which can easily be converted into a matrix or a data frame (which is the input format for `ExomeDepth`). An example of `GenomicRanges` output generated by `getBAMCounts` is provided in this package (chromosome 1 only to keep the size manageable). Here is how this object could for example be used to obtain a more generic data frame:

```
> library(ExomeDepth)
> data(ExomeCount)
> ExomeCount.dafr <- as(ExomeCount[, colnames(ExomeCount)], 'data.frame')
> ExomeCount.dafr$chromosome <- gsub(as.character(ExomeCount.dafr$space),
+                                   pattern = 'chr',
+                                   replacement = '') ##remove the annoying chr letters
> print(head(ExomeCount.dafr))
```

	space	start	end	width	names	GC	Exome1	Exome2	Exome3	Exome4
1	1	12012	12058	47	DDX11L10-201_1	0.6170213	0	0	0	0
2	1	12181	12228	48	DDX11L10-201_2	0.5000000	0	0	0	0
3	1	12615	12698	84	DDX11L10-201_3	0.5952381	118	242	116	170
4	1	12977	13053	77	DDX11L10-201_4	0.6103896	198	48	104	118
5	1	13223	13375	153	DDX11L10-201_5	0.5882353	516	1112	530	682
6	1	13455	13671	217	DDX11L10-201_6	0.5898618	272	762	336	372

```

chromosome
1          1
2          1
3          1
4          1
5          1
6          1

```

2.2 Counts for chromosome X

Calling CNVs on the X chromosome can create issues if the exome sample of interest and the reference exome samples it is being compared to (what I call the aggregate reference) are not gender matched. For this reason the chromosome X exonic regions are not included by default in the data frame `exons.hg19`, mostly to avoid users getting low quality CNV calls because of this issue. However, loading the same dataset in R also brings another object called `exons.hg19.X` that contains the chromosome X exons.

```

> data(exons.hg19.X)
> head(exons.hg19.X)

```

```

      chromosome  start    end      name
238055          X 170411 170514 NCRNA00108-001_1
238056          X 171605 171759 NCRNA00108-001_2
238057          X 172683 172713 NCRNA00108-001_3
238058          X 192992 193062   PLCXD1-201_1
238059          X 198150 198352   PLCXD1-201_2
238060          X 200835 200982   PLCXD1-201_3

```

This object can be used to generate CNV counts and further down the line CNV calls, in the same way as `exons.hg19`. While this is not really necessary, I would recommend calling CNV on the X separately from the autosomes. Also make sure that the genders are matched properly (i.e. do not use male as a reference for female samples and vice versa).

3 Load an example dataset

We have already loaded a dataset of chromosome 1 data for four exome samples. We run a first test to make sure that the model can be fitted properly. Note the use of the `subset.for.speed` option that subsets some rows purely to speed up this computation.

```

> test <- new('ExomeDepth',
+           test = ExomeCount.dafr$Exome2,
+           reference = ExomeCount.dafr$Exome3,
+           formula = 'cbind(test, reference) ~ 1',
+           subset.for.speed = seq(1, nrow(ExomeCount.dafr), 100))
> show(test)

```

Number of data points: 266

Formula: `cbind(test, reference) ~ 1`

Phi parameter (range if multiple values have been set): 0.0229405 0.0229405

Likelihood computed

4 Build the most appropriate reference set

A key idea behind ExomeDepth is that each exome should not be compared to all other exomes but rather to an optimized set of exomes that are well correlated with that exome. This is what I call the optimized aggregate reference set, which is optimized for each exome sample. So the first step is to select the most appropriate reference sample. This step is demonstrated below.

```

> my.test <- ExomeCount$Exome4
> my.ref.samples <- c('Exome1', 'Exome2', 'Exome3')
> my.reference.set <- as.matrix(ExomeCount.dafr[, my.ref.samples])
> my.choice <- select.reference.set (test.counts = my.test,
+                                   reference.counts = my.reference.set,
+                                   bin.length = (ExomeCount.dafr$end - ExomeCount.dafr$start)/1000,
+                                   n.bins.reduced = 10000)
> print(my.choice[[1]])

[1] "Exome2" "Exome1" "Exome3"

```

Using the output of this procedure we can construct the reference set.

```

> my.reference.selected <- apply(X = as.matrix( ExomeCount.dafr[, my.choice$reference.choice] ),
+                               MAR = 1,
+                               FUN = sum)

```

5 CNV calling

Now the following step is the longest one as the beta-binomial model is applied to the full set of exons:

```

> all.exons <- new('ExomeDepth',
+                 test = my.test,
+                 reference = my.reference.selected,
+                 formula = 'cbind(test, reference) ~ 1')

```

We can now call the CNV by running the underlying hidden Markov model:

```

> all.exons <- CallCNVs(x = all.exons,
+                       transition.probability = 10^-4,
+                       chromosome = ExomeCount.dafr$space,
+                       start = ExomeCount.dafr$start,
+                       end = ExomeCount.dafr$end,
+                       name = ExomeCount.dafr$names)

```

Number of hidden states: 3

Number of data points: 26547

Initializing the HMM

Done with the first step of the HMM, now running the trace back

Total number of calls: 23

```

> print(head(all.exons@CNV.calls))

```

	start.p	end.p	type	nexons	start	end	chromosome
1	25	27	deletion	3	89553	91106	1
2	52	66	deletion	15	324290	523834	1
3	100	103	duplication	4	743956	745551	1
4	575	576	deletion	2	1569583	1570002	1
5	587	591	deletion	5	1592941	1603069	1
6	2324	2327	deletion	4	12976452	12980570	1

	id	BF	reads.expected	reads.observed	reads.ratio
1	chr1:89553-91106	12.40	224	68	0.304
2	chr1:324290-523834	13.40	380	190	0.500
3	chr1:743956-745551	7.67	201	336	1.670
4	chr1:1569583-1570002	5.53	68	24	0.353
5	chr1:1592941-1603069	13.90	1136	434	0.382
6	chr1:12976452-12980570	12.10	780	342	0.438

Now the last thing to do is to save it in an easily readable format (csv in this example, which can be opened in excel if needed):

```
> output.file <- 'exome_calls.csv'
> write.csv(file = output.file,
+           x = all.exons@CNV.calls,
+           row.names = FALSE)
```

Note that it is probably best to annotate the calls before creating that csv file (see below for annotation tools).

6 Better annotation of CNV calls

Much can be done to annotate CNV calls and this is a difficult problem. While this is a work in progress, I have started adding basic options. The first, and perhaps the most useful, is to identify the overlap with a set of common CNVs identified in Conrad et al, Nature 2010. If one is looking for rare CNVs, these should probably be filtered out. The first step is to load these reference data from Conrad et al. To make things as practical as possible, these data are now available as part of ExomeDepth.

```
> data(Conrad.hg19)
> head(Conrad.hg19.common.CNVs)
```

GRanges with 6 ranges and 1 elementMetadata col:

	seqnames	ranges	strand	names
	<Rle>	<IRanges>	<Rle>	<factor>
[1]	1	[10499, 91591]	*	CNVR1.1
[2]	1	[10499, 177368]	*	CNVR1.2
[3]	1	[82705, 92162]	*	CNVR1.5
[4]	1	[85841, 91967]	*	CNVR1.4
[5]	1	[87433, 89163]	*	CNVR1.6
[6]	1	[87446, 109121]	*	CNVR1.7

seqlengths:

	1	10	11	12	13	14	15	16	17	18	19	2	20	21	22	3	4	5	6	7	8	9	X
NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA

Then one can use this information to annotate our CNV calls with the function `AnnotateExtra`.

```
> all.exons <- AnnotateExtra(x = all.exons,
+                             reference.annotation = Conrad.hg19.common.CNVs,
+                             min.overlap = 0.5,
+                             column.name = 'Conrad.hg19')
```

The `min.overlap` argument set to 0.5 requires that the Conrad reference call overlaps at least 50% of our CNV calls to declare an overlap. The `column.name` argument simply defines the name of the column that will store the overlap information. The outcome of this procedure can be checked with:

```
> print(head(all.exons@CNV.calls))
```

	start.p	end.p	type	nexons	start	end	chromosome
1	25	27	deletion	3	89553	91106	1
2	52	66	deletion	15	324290	523834	1
3	100	103	duplication	4	743956	745551	1
4	575	576	deletion	2	1569583	1570002	1
5	587	591	deletion	5	1592941	1603069	1
6	2324	2327	deletion	4	12976452	12980570	1

	id	BF	reads.expected	reads.observed	reads.ratio
1	chr1:89553-91106	12.40	224	68	0.304
2	chr1:324290-523834	13.40	380	190	0.500
3	chr1:743956-745551	7.67	201	336	1.670
4	chr1:1569583-1570002	5.53	68	24	0.353
5	chr1:1592941-1603069	13.90	1136	434	0.382

```

6 chr1:12976452-12980570 12.10          780          342          0.438
      Conrad.hg19
1 CNVR1.1,CNVR1.2,CNVR1.5,CNVR1.4,CNVR1.7
2      CNVR2.4
3      <NA>
4      CNVR17.1
5      CNVR17.1
6      CNVR72.3,CNVR72.4,CNVR72.2

```

I have processed the Conrad et al data in the GRanges format. Potentially any other reference dataset could be converted as well. See for example the exon information:

```

> exons.hg19.GRanges <- GRanges(seqnames = exons.hg19$chromosome,
+                               IRanges(start=exons.hg19$start,end=exons.hg19$end),
+                               names = exons.hg19$name)
> all.exons <- AnnotateExtra(x = all.exons,
+                             reference.annotation = exons.hg19.GRanges,
+                             min.overlap = 0.0001,
+                             column.name = 'exons.hg19')
> all.exons@CNV.calls[3:6,]

```

	start.p	end.p	type	nexons	start	end	chromosome
3	100	103	duplication	4	743956	745551	1
4	575	576	deletion	2	1569583	1570002	1
5	587	591	deletion	5	1592941	1603069	1
6	2324	2327	deletion	4	12976452	12980570	1

	id	BF	reads.expected	reads.observed	reads.ratio
3	chr1:743956-745551	7.67	201	336	1.670
4	chr1:1569583-1570002	5.53	68	24	0.353
5	chr1:1592941-1603069	13.90	1136	434	0.382
6	chr1:12976452-12980570	12.10	780	342	0.438

```

      Conrad.hg19
3      <NA>
4      CNVR17.1
5      CNVR17.1
6 CNVR72.3,CNVR72.4,CNVR72.2

```

```

      exons.hg19
3      ENST00000447500_1,ENST00000447500_2,ENST00000412115_3,ENST00000435300_2
4      MMP23B-202_8,MMP23B-202_9
5 RP11-345P4-003_6,RP11-345P4-003_7,RP11-345P4-003_8,RP11-345P4-003_9,RP11-345P4-003_10
6      PRAMEF7-201_1,PRAMEF7-201_2,PRAMEF7-201_3,PRAMEF7-201_4

```

This time I report any overlap with an exon by specifying a value close to 0 in the `min.overlap` argument. Note the metadata column `names` which MUST be specified for the annotation to work properly.

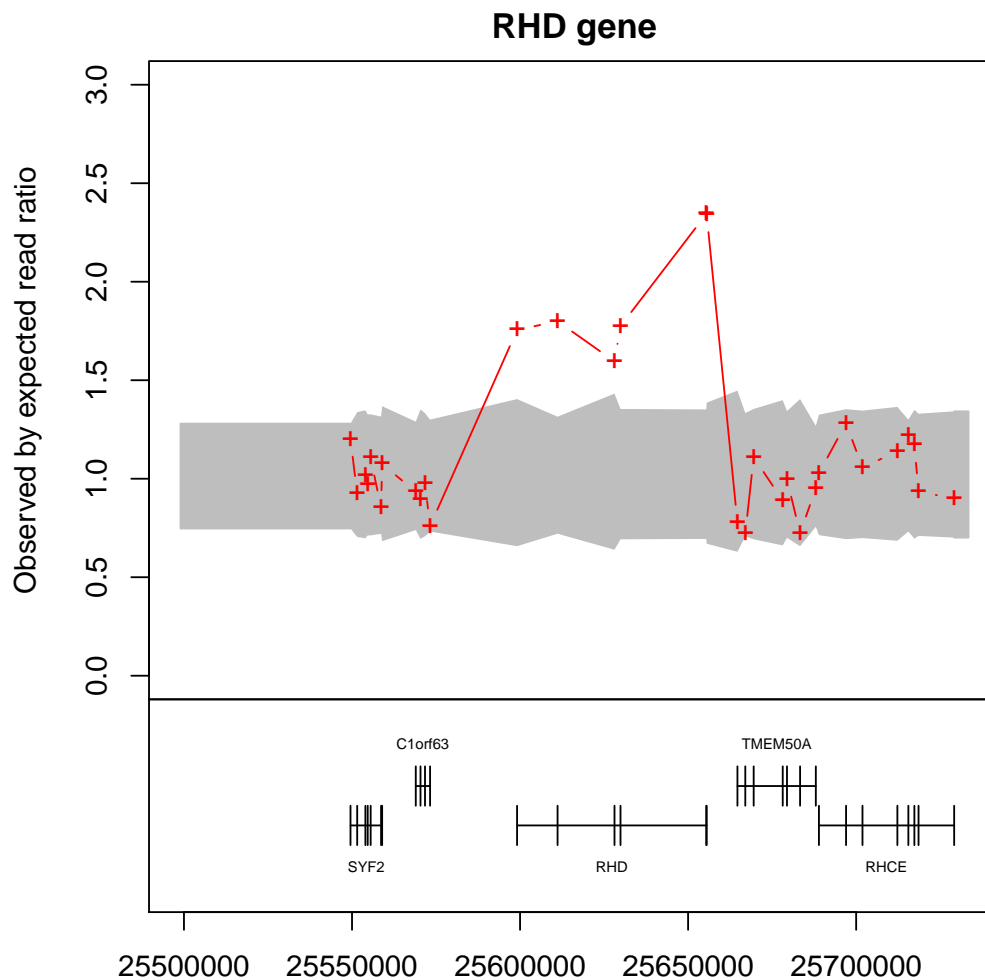
7 A visual example

The ExomeDepth object includes a plot function. This function shows the ratio between observed and expected read depth. The 95% confidence interval is marked by a grey shaded area. Here we use a common CNV located in the RHD gene as an example. We can see that the individual in question has more copies than the average (in fact two functional copies of RHD, which corresponds to rhesus positive).

```

> plot (all.exons,
+       sequence = '1',
+       xlim = c(25598981 - 100000, 25633433 + 100000),
+       count.threshold = 20,
+       main = 'RHD gene',
+       with.gene = TRUE)

```



8 How to loop over the multiple samples

A FAQ is a way to deal with a set of a significant number of exomes, i.e. how to loop over all of them using ExomeDepth. This can be done with a loop. I show below an example of how I would code things. The code is not executed in the vignette to save time when building the package, but it can give some hints to users who do not have extensive experience with R.

```
> ExomeCount.mat <- as.matrix(ExomeCount.dafr[, grep(names(ExomeCount.dafr), pattern = 'Exome.*')])
> nsamples <- ncol(ExomeCount.mat)
> for (i in 1:nsamples) {
+
+ ##### Create the aggregate reference set for this sample
+ my.choice <- select.reference.set (test.counts = ExomeCount.mat[,i],
+                                   reference.counts = ExomeCount.mat[,-i],
+                                   bin.length = (ExomeCount.dafr$end - ExomeCount.dafr$start)/1000,
+                                   n.bins.reduced = 10000)
+
+ my.reference.selected <- apply(X = ExomeCount.mat[, my.choice$reference.choice],
+                               MAR = 1,
+                               FUN = sum)
+
+ message('Now creating the ExomeDepth object')
```

```

+ all.exons <- new('ExomeDepth',
+               test = ExomeCount.mat[,i],
+               reference = my.reference.selected,
+               formula = 'cbind(test, reference) ~ 1')
+
+ ##### Now call the CNVs
+ all.exons <- CallCNVs(x = all.exons,
+                      transition.probability = 10^-4,
+                      chromosome = ExomeCount.dafr$space,
+                      start = ExomeCount.dafr$start,
+                      end = ExomeCount.dafr$end,
+                      name = ExomeCount.dafr$names)
+
+ ##### Now annotate the ExomeDepth object
+ all.exons <- AnnotateExtra(x = all.exons,
+                           reference.annotation = Conrad.hg19.common.CNVs,
+                           min.overlap = 0.5,
+                           column.name = 'Conrad.hg19')
+
+ all.exons <- AnnotateExtra(x = all.exons,
+                           reference.annotation = exons.hg19.GRanges,
+                           min.overlap = 0.0001,
+                           column.name = 'exons.hg19')
+
+ output.file <- paste('Exome_', i, 'csv', sep = '')
+ write.csv(file = output.file, x = all.exons@CNV.calls, row.names = FALSE)
+
+ }
+
+
+

```

9 Technical information about R session

```
> sessionInfo()
```

```
R version 2.15.1 (2012-06-22)
```

```
Platform: x86_64-unknown-linux-gnu (64-bit)
```

```
locale:
```

```

[1] LC_CTYPE=en_US.iso885915      LC_NUMERIC=C
[3] LC_TIME=en_US.iso885915      LC_COLLATE=C
[5] LC_MONETARY=en_US.iso885915  LC_MESSAGES=en_US.iso885915
[7] LC_PAPER=C                    LC_NAME=C
[9] LC_ADDRESS=C                  LC_TELEPHONE=C
[11] LC_MEASUREMENT=en_US.iso885915 LC_IDENTIFICATION=C

```

```
attached base packages:
```

```

[1] stats4      splines      stats        graphics     grDevices    utils        datasets
[8] methods     base

```

```
other attached packages:
```

```

[1] ExomeDepth_0.9.1      Rsamtools_1.8.5      Biostrings_2.24.1
[4] GenomicRanges_1.8.11 IRanges_1.14.4       BiocGenerics_0.2.0
[7] VGAM_0.8-7            aod_1.3

```

```
loaded via a namespace (and not attached):
```

```
[1] bitops_1.0-4.1 tools_2.15.1  zlibbioc_1.2.0
```